
Published version (with publisher's formatting)

This version is available at: https://eprints.mdx.ac.uk/792/

Copyright:

Middlesex University Research Repository makes the University's research available electronically.

Copyright and moral rights to this work are retained by the author and/or other copyright owners unless otherwise stated. The work is supplied on the understanding that any use for commercial gain is strictly forbidden. A copy may be downloaded for personal, non-commercial, research or study without prior permission and without charge.

Works, including theses and research projects, may not be reproduced in any format or medium, or extensive quotations taken from them, or their content changed in any way, without first obtaining permission in writing from the copyright holder(s). They may not be sold or exploited commercially in any format or medium without the prior written permission of the copyright holder(s).

Full bibliographic details must be given when referring to, or quoting from full items including the author’s name, the title of the work, publication details where relevant (place, publisher, date), pagination, and for theses or dissertations the awarding institution, the degree type awarded, and the date of the award.

If you believe that any material held in the repository infringes copyright law, please contact the Repository Team at Middlesex University via the following email address:

eprints@mdx.ac.uk

The item will be removed from the repository while any claim is being investigated.

See also repository copyright: re-use policy: http://eprints.mdx.ac.uk/policies.html#copy
The temporary anatomical structures prominent in the first trimester may be fulfilling exchange functions assigned to the placenta in the second and third trimester

S.M. Docherty, R.K. Iles, N. Wathen and T. Chard

Departments of Obstetrics, Gynaecology and Reproductive Physiology, St. Bartholomew's Hospital Medical College, West Smithfield, London EC1A 7BE, UK

Introduction

The introduction of high-resolution transvaginal ultrasonography has allowed observation and sampling of the embryonic cavities of early gestation (Jauniaux et al., 1991b; Wathen et al., 1991a). This has given new insights into the composition of extra-embryonic coelomic (EEC) fluid and amniotic fluid in the first trimester of pregnancy, and has generated novel hypotheses on the physiology and biochemistry of early gestation.

Formation of the extra-embryonic coelom and yolk sac

After fertilization, a series of rapid mitoses in the zygote forms the morula, which reaches the endometrial cavity 4–5 days later. Accumulation of fluid in intercellular spaces leads to formation of the blastocyst; this comprises an outer layer which will form the trophoblast, an inner cell mass and the blastocoelic fluid, all still encased within the zona pellucida. Implantation commences 7–8 days post-ovulation. The disappearance of the zona pellucida exposes the trophoblast, which adheres to the epithelial cells of the endometrium, penetrates this layer and invades the underlying stroma. This leads to the ‘decidual reaction’; the endometrial stroma surrounding the implantation site becomes highly oedematous and vascular, with appearance of large granular lymphocytes.

At the time of implantation the cells of the inner cell mass differentiate to form two layers: a hypoblast layer adjacent to the blastocyst cavity, and an epiblast layer. Together the hypoblast and epiblast form the bilaminar germ disk. The amniotic cavity forms between the epiblast and the trophoblast, while some of the most caudal cells of the epiblast layer proliferate to form a thin membrane which lines the inner surface of the trophoblastic basal lamina and forms the primitive yolk sac. A new population of cells, the extra-embryonic mesoderm, evolves and separates the trophoblast and the yolk sac. Cavities form in this layer which coalesce to form the EEC (Carlson, 1994). The coelom divides the extra-embryonic mesoderm into two layers. Somatic mesoderm lines the trophoblast, and splanchnic mesoderm surrounds the secondary yolk sac and fetus. Hypoblastic cells migrating from the embryonic disk along the inside of the exocoelomic membrane proliferate and lead to formation of the much smaller secondary yolk sac. During the third week of gestation, gastrulation establishes all three germ layers in the embryo. The primitive streak forms on the surface of the epiblast, with the primitive node at its cephalic end. Epiblast cells invaginate in the region of the streak to form the endoderm and mesoderm. The secondary yolk sac communicates with the ventral aspect of the trilaminar embryo, while the cytotrophoblast-derived amnion meets the ventral aspect of the developing embryo at the site of umbilical cord insertion. As the embryo enlarges, so the amniotic cavity around it expands relative to the EEC. Lateral folding of the embryo during the sixth week of gestation results in narrowing of the base of the yolk sac, with the formation of the yolk stalk. The yolk sac itself becomes more peripheral as gestation progresses. The structures present at around the ninth week of gestation are shown in Figure 1.

Fate of the extra-embryonic coelom and yolk sac

By the 13th week of gestation the amniotic cavity has expanded so that the amnion is in direct contact with the trophoblast; thus the EEC is obliterated. The yolk sac becomes increasingly marginalized and the yolk stalk reduced to a narrow vitelline duct as the amniotic cavity expands. The yolk sac degenerates at around the same time as its stalk becomes covered in amnion and incorporated into the primitive umbilical cord. The connection with the midgut via the vitelline duct is also obliterated at this time (Sadler, 1990).

Possible functions of the extra-embryonic coelom

Recent studies have shown that the composition of the coelomic fluid differs dramatically from that of amniotic fluid, and that there are great variations with gestational age (Jauniaux et al.,...
membrane of the amnion (King, 1985) may result in electrostatic repulsion and steric hindrance of large anionic molecules, and thus represent a significant barrier to permeability (Jones and Jauniaux, 1995). However, a low permeability to anionic, large molecules would not explain the disparity in the amounts of the low molecular weight, lipophilic steroids and vitamins (Campbell et al., 1993; Jauniaux et al., 1993; Campbell et al., 1994; Sourial et al., 1994), nor the equal distribution of AFP. Active transport mechanisms in a dynamic amnion might, therefore, be responsible for initiating and maintaining the disparities. This concept is supported by the presence of high concentrations of bicarbonate in the amniotic fluid and high concentrations of phosphate in the EEC, suggesting the presence of ion anti-ports driving active transport channels. Concentrations of some of these ions in amniotic and coelomic fluids are summarized in Table II. A number of important changes have been observed in the ultrastructure of the amnion during the first trimester. Rich stores of intracellular glycogen are present in the amniotic epithelium during early pregnancy and may represent an energy source in the absence of organelles such as mitochondria or endoplasmic reticulum (Jones and Jauniaux, 1995). The amnion has carbonic anhydrase activity (Benirschke and Kaufmann, 1990). Carbonic anhydrase is an enzyme which catalyses the formation of carbonic acid from carbon dioxide and water, then allowing the dissociation of carbonic acid to bicarbonate and hydrogen ions. This may imply a role for the amnion in the regulation of pH in amniotic fluid (Jones and Jauniaux, 1995). Similarly, 17β-hydroxysteroid dehydrogenase activity has been reported in the amnion between 7–20 weeks of gestation (Sulcová et al., 1974), which may suggest a role for amnion epithelial cells in the modification of steroids. One possible alternative explanation for the function of the amnion may be as an

### Table I. Median value of pregnancy-associated antigens in amniotic fluid, extra-embryonic coelomic (EEC) fluid and maternal serum between 8–12 weeks gestation*

<table>
<thead>
<tr>
<th>Pregnancy-associated antigen</th>
<th>Reference</th>
<th>Tissue source</th>
<th>Amniotic fluid</th>
<th>EEC fluid</th>
<th>Maternal serum</th>
</tr>
</thead>
<tbody>
<tr>
<td>Alpha fetoprotein (kIU/ml)</td>
<td>Wathen et al., 1992b</td>
<td>Fetal yolk sac</td>
<td>26.0</td>
<td>24.1</td>
<td>6.4</td>
</tr>
<tr>
<td>Alpha fetoprotein (kIU/ml)</td>
<td>Jauniaux et al., 1993</td>
<td>Fetal yolk sac</td>
<td>6.1</td>
<td>7.1</td>
<td>-</td>
</tr>
<tr>
<td>Cancer antigen 125 (IU/ml)</td>
<td>Campbell et al., 1992a</td>
<td>Fetal</td>
<td>496</td>
<td>35</td>
<td>35</td>
</tr>
<tr>
<td>HCG (IU/ml)</td>
<td>Iles et al., 1992</td>
<td>Placental trophoblast</td>
<td>1.73</td>
<td>245</td>
<td>157</td>
</tr>
<tr>
<td>HCG (IU/ml)</td>
<td>Jauniaux et al., 1993</td>
<td>Placental trophoblast</td>
<td>1.00</td>
<td>120</td>
<td>81</td>
</tr>
<tr>
<td>Total β-subunit HCG (IU/ml)</td>
<td>Iles et al., 1992</td>
<td>Placental trophoblast</td>
<td>6.73</td>
<td>410</td>
<td>141.5</td>
</tr>
<tr>
<td>Free α-subunit HCG (μg/ml)</td>
<td>Iles et al., 1992</td>
<td>Placental trophoblast</td>
<td>0.262</td>
<td>17.3</td>
<td>1.3</td>
</tr>
<tr>
<td>Progesterone (pmol/ml)</td>
<td>Unpublished data</td>
<td>Placental trophoblast</td>
<td>56.0</td>
<td>850</td>
<td>83.5</td>
</tr>
<tr>
<td>Progesterone (pmol/ml)</td>
<td>Jauniaux et al., 1993</td>
<td>Placental trophoblast</td>
<td>23.6</td>
<td>877</td>
<td>69.7</td>
</tr>
<tr>
<td>Oestradiol (pmol/ml)</td>
<td>Jauniaux et al., 1993</td>
<td>Placental trophoblast</td>
<td>5083</td>
<td>26978</td>
<td>4448</td>
</tr>
<tr>
<td>Unconjugated oestradiol (pmol/ml)</td>
<td>Wathen et al., 1992a</td>
<td>Placental trophoblast</td>
<td>&lt;1.2</td>
<td>2.6</td>
<td>&lt;1.2</td>
</tr>
<tr>
<td>Placental protein 14 (ng/ml)</td>
<td>Wathen et al., 1992a</td>
<td>Placental trophoblast</td>
<td>77</td>
<td>4416</td>
<td>642</td>
</tr>
<tr>
<td>IGFBP-1 (ng/ml)</td>
<td>Wathen et al., 1992b</td>
<td>Fetal/Maternal</td>
<td>7.5</td>
<td>500</td>
<td>64</td>
</tr>
<tr>
<td>Inulin-like growth factor-1 (ng/ml)</td>
<td>Wathen et al., 1992b</td>
<td>Fetal/Maternal</td>
<td>273</td>
<td>364</td>
<td>98</td>
</tr>
<tr>
<td>Human placental lactogen (ng/ml)</td>
<td>Wathen et al., 1992a</td>
<td>Placental syncytiotrophoblast</td>
<td>30.0</td>
<td>80.0</td>
<td>210.0</td>
</tr>
<tr>
<td>Prolactin (IU/ml)</td>
<td>Wathen et al., 1993</td>
<td>Maternal decidua</td>
<td>40</td>
<td>371</td>
<td>709</td>
</tr>
<tr>
<td>Vitamin A (μmol/l)</td>
<td>Campbell et al., 1994</td>
<td>-</td>
<td>0</td>
<td>0.08</td>
<td>1.85</td>
</tr>
<tr>
<td>Vitamin E (μmol/l)</td>
<td>Campbell et al., 1994</td>
<td>-</td>
<td>0</td>
<td>0.26</td>
<td>17.01</td>
</tr>
<tr>
<td>Folate (μg/l)</td>
<td>Campbell et al., 1993</td>
<td>-</td>
<td>2.15</td>
<td>9.9</td>
<td>6.2</td>
</tr>
<tr>
<td>Vitamin B12 (ng/l)</td>
<td>Campbell et al., 1993</td>
<td>-</td>
<td>987</td>
<td>3680</td>
<td>405</td>
</tr>
<tr>
<td>Cobalamin (ng/l)</td>
<td>Sourial et al., 1994</td>
<td>-</td>
<td>589</td>
<td>3162</td>
<td>427</td>
</tr>
<tr>
<td>Pregnancy-associated plasma protein A (mIU/ml)</td>
<td>Iles et al., 1994</td>
<td>Placental syncytiotrophoblast</td>
<td>&lt;10</td>
<td>26</td>
<td>1220</td>
</tr>
</tbody>
</table>

HCG = human chorionic gonadotrophin; IGFBP-1 = insulin-like growth factor binding protein-1.
Figure 2. Median concentrations of placental protein-14 (PP14) and albumin in amniotic fluid from 10-20 weeks gestation (taken from Chatzakis et al., 1994).

Figure 3. Concentrations of prolactin and insulin-like growth factor binding protein-1 (IGFBP-1) in amniotic fluid from 9-20 weeks gestation (taken from Wathen et al., 1993).

Table II. Biochemical composition* of amniotic and coelomic fluid as measured at 7-12 weeks gestation in normal pregnancy and analysed using Student's t-test

<table>
<thead>
<tr>
<th>Variable</th>
<th>Amniotic fluid</th>
<th>Coelomic fluid</th>
<th>Significance (P)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sodium (mmol/l)</td>
<td>141.2</td>
<td>138.1</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td>Potassium (mmol/l)</td>
<td>3.98</td>
<td>3.86</td>
<td>0.0038</td>
</tr>
<tr>
<td>Chloride (mmol/l)</td>
<td>97.41</td>
<td>110.1</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td>Urea (mmol/l)</td>
<td>3.32</td>
<td>3.9</td>
<td>0.02</td>
</tr>
<tr>
<td>Bicarbonate (mmol/l)</td>
<td>34.24</td>
<td>20.9</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td>Creatinine (µmol/l)</td>
<td>37.1</td>
<td>72.31</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td>Calcium (mmol/l)</td>
<td>1.43</td>
<td>2.66</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td>Phosphate (µmol/l)</td>
<td>1.09</td>
<td>2.69</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td>Bilirubin (µmol/l)</td>
<td>0.64</td>
<td>3.4</td>
<td>&lt;0.0001</td>
</tr>
</tbody>
</table>

*From Campbell et al., 1992b.

Exchange functions in the first trimester

A prerequisite for induction of placental gene expression (Rodesch et al., 1992). This has been demonstrated for transcription of the gene for vascular endothelial growth factor (VEGF) which is increased in placental fibroblasts grown in hypoxic conditions (Wheeler et al., 1995). The expansion and obliteration of the EEC also coincides with the rise and fall in concentrations of HCG in the first trimester, leading to the suggestion that a tissue closely associated with the EEC (probably the chorion) is the principle source of HCG (Chard et al., 1995). It is notable that concentrations of HCG and its subunits are strikingly higher in the EEC than elsewhere, including maternal blood (Iles et al., 1992; Nagy et al., 1994; Jauniaux et al., 1995).

The EEC may also be involved in the delivery of nutrients to the fetus via the secondary yolk sac and vitelline circulation. Vitamins A, B12, E, folate and cobalamin are present in high concentrations in the EEC (Table I) (Campbell et al., 1993, 1994; Sourial et al., 1994)

AFP is unusual in that concentrations are equivalent in both amniotic and exocoelomic cavities; the molecular variants found in both sites show a common derivation from the yolk sac (Jauniaux et al., 1993). AFP is the fetal analogue of albumin (Alpert et al., 1971); there is a 39% sequence homology with the adult molecule (Morinaga et al., 1983), and both are coded on the long arm of chromosome 4. AFP may function similarly to its adult equivalent in showing binding affinities for a variety of ligands. However, AFP differs from albumin in its preferential binding of polyunsaturated fatty acids (reviewed by Deutsch, 1991) and may have a role in transporting these substances to developing cells; AFP could be a transport protein for nutrients and growth factors on both sides of the amnion.

Possible functions of the yolk sac

The secondary yolk sac floats in the EEC, and the cavity is in open connection with the midgut of the developing fetus via the vitelline duct. Its wall comprises an external mesothelial layer of flattened cells (facing the EEC), a vascular mesenchyme, and an endodermal layer of columnar cells facing into the yolk sac cavity (Jauniaux and Moscoso, 1992). It has a well developed microvillous border on its external surface and numerous pinocytotic vesicles, implying an active absorptive function extremely effective barrier against waste products accumulated in the EEC via an as yet unidentified route.

The EEC may segregate highly active molecules including differentiation factors, cytokines, hormones and waste products, thus protecting the poorly keratinized (i.e. highly permeable) fetus at a time when crucial differentiation and organogenesis is occurring. After the obliteration of the EEC at around 12 weeks of gestation, there is a dramatic rise in the concentration of most placental and endometrial proteins in the amniotic fluid (Wathen et al., 1993; Chatzakis et al., 1994; Iles et al., 1994), for example, PP-14 (Figure 2), prolactin and IGFBP-1 (Figure 3). This striking discontinuity at 12–14 weeks is consistent with the loss of transport function by the amnion, since fusion of the amnion and trophoblast would eliminate the reservoir (i.e. the EEC) for export of such proteins. These important changes in transport functions coincide with the appearance of maternal blood flow in the intervillous space (Hustin and Schaaps, 1987).

Removal of waste products from the proximity of the sensitive embryo via the EEC could be an important feature of early gestation. Only later, with the onset of maternal blood flow in the intervillous space, does the placenta subsume this function. Indeed, a state of relative hypoxia is now considered...
function (Jauniaux et al., 1991a) in addition to its documented biosynthetic functions. The yolk sac produces AFP in its endodermal layer until the 10th week of gestation (Gitlin and Perricelli, 1970). It is probably the major source of protein synthesis (Gulbis et al., 1992) (including albumin and ferritin) at a time prior to embryonic liver maturation. Protein synthesis by the yolk sac ceases after the ninth week of gestation (Jones and Jauniaux, 1995). In addition, the yolk sac synthesizes many enzymes involved in digestion and metabolism, including lactate dehydrogenase, galactosidase, α-glutamyl transferase and acid phosphatase (Buffe et al., 1993). The yolk sac could represent a source of nutrients to the growing embryo at a time when there is relatively poor placental blood supply, and it might also absorb waste products from the EEC, using these in the manufacture of essential molecules. The yolk sac could thus be described as a primitive extra-embryonic 'liver' to the early embryo.

The yolk sac is the first site of haematoopoiesis (Moore and Metcalf, 1970), producing nucleated red blood cells in 'blood islands' and seeding the fetal liver and spleen for further haematoopoiesis after its decline (reviewed by Tavassoli, 1991), though the precise site of initiation is still unknown. The yolk sac is also proposed as the source of primordial germ cells (Witschi, 1948). Nothing more is known of the functions of the human yolk sac, but the rodent yolk sac is more amenable to investigation; functions include nutritional, endocrine, secretory, excretory, metabolic, haematogenic and immunological (reviewed by Beckman et al., 1990). Furthermore, agents which target the rat yolk sac are highly embryotoxic, emphasizing the overall physiological importance of this structure (Beckman et al., 1990; Brent et al., 1990; Jollie, 1990).

In humans, a possible link between a small or absent yolk sac and spontaneous abortion has been suggested (Nogales et al., 1993)

Conclusion

The fact that there are high concentrations of proteins, steroids, and other organic and inorganic molecules in the coelomic fluid but not the amniotic fluid in the first trimester, together with the absence of inter villous blood flow at that time, suggests a pivotal role for the amnion and EEC in segregation and excretion of active molecules and waste products from the developing embryo. The site and prominence of the yolk sac suggests a crucial role in nutritional, endocrine and metabolic support of the early conceptus.

The disappearance of the EEC and the secondary yolk sac at around 12 weeks of pregnancy coincides with an abrupt increase in amniotic fluid levels of various biological molecules, and the start of perfusion of the inter villous space by maternal blood. This implies that the developed placenta takes over the functions previously fulfilled by the yolk sac, amnion and EEC.

References


Exchange functions in the first trimester


Received on December 28, 1995; accepted on March 21, 1996